[©]ISISnet Publishers

Seasonal variation of microsporidiosis among HIV-infected persons in Benin City, Nigeria.

F.O. Akinbo¹, C. E. Okaka², R. Omoregie³, O.Igbinuwen⁴ and C. Egbe⁵

¹Department of Pathology, University of Benin Teaching Hospital, Benin City, **Nigeria**.

²Department of Animal and Environmental Biology, University of Benin, Benin City, **Nigeria**.

³School of Medical Laboratory sciences, University of Benin Teaching Hospital, Benin City, **Nigeria**.

⁴Department of Haematology, University of Benin Teaching Hospital, Benin City, **Nigeria**.

⁵Department of Medical Microbiology, University of Benin Teaching Hospital, Benin City, **Nigeria**.

*Corresponding author

The prevalence of microsporidiosis in relation to seasonal variations is unknown. This study is aimed at determining the correlation between seasonal variation and the prevalence of microsporidiosis in HIV-infected persons in Benin City, Nigeria. A total of 763 persons consisting of 463 HIV-infected patients attending HIV clinics and 300 apparently healthy HIV non-infected persons were recruited in this study. Stool specimens were collected from each patient and processed using standard procedure. The prevalence of microsporidiosis was significantly associated with season. More episodes of intestinal parasitic infections were observed in the rainy season than the dry season (OR = 1.990; 95% CI = 1.374, 2.882; p = 0.0004). This study underscores the association between microsporidiosis and seasonal variations with higher prevalence during the rainy season.

Key words: Benin City, HIV, microsporidiosis, Seasonal variation.

Microsporidia species is one of the common causes of chronic diarrhea in AIDS patients (van Gool et al., 1995; Tumwine et al., 2002) which is responsible for unexplained weight loss in HIV- infected persons (Sokolova et al., 2011). The clinical manifestations of microsporidiosis are diverse and include intestinal, pulmonary, ocular, muscular, and renal disease (Didier and Weiss, 2006). The correlation between seasonal variation and the prevalence of microsporidiosis is still under investigation. A study in the United States (Conteas et al., 1998) and another in Brazil (Cotte et al., 1999) did not observe seasonal trend in the prevalence of microsporidiosis. Data are lacking on the association between seasonal variation and the prevalence of microsporidiosis in HIVinfected persons in Nigeria. This study is

aimed at determining the correlation between seasonal variation and the prevalence of microsporidiosis in HIV-infected persons in Benin City, Nigeria.

MATERIALS AND METHODS Study population

The study was carried out at the University of Benin Teaching Hospital, Benin City – a teaching hospital with a referral status and center for HIV/AIDS management under the United States President's Emergency Plan for AIDS Relief (PEPFAR). A total of 763 persons consisting of 463 HIV-infected patients attending HIV clinics and 300 apparently healthy HIV non-infected individuals that were contacted through HIV outreach programs in their homes and offices that served as controls were included in this study. Individuals that were on highly active antiretroviral therapy (HAART), antiparasitic agents and those with AIDS defining conditions were excluded from this study. Verbal informed consent was obtained from each participant. This study was approved by the Ethical Committee of the University of Benin Teaching Hospital, Benin City.

Specimen collection and processing

Stool specimens were collected from each participant into clean wide-mouthed container. The freshly voided stool specimens processed using formol-ether were examined concentration method and microscopically for intestinal parasites as previously described (Cheesbrough, 1999). Briefly, about 1g of faeces was emulsified in 4ml of formol saline and agitated. The mixture was sieved. To the filtrate, 4ml of diethyl ether was added and mixed. The filtrate-ether mixture was spun at 3,000rpm for 1 minute. The faecal debris on the side of the tube was detached with the aid of a plastic pipette and supernatant discarded. From the this sediment, saline and iodine mounts were prepared and examined for presence of parasites. Following this, each fresh stool sample was preserved in 10% formol saline. From this, a concentrated smear was made on glass slide and stained by Ryan's modified Trichrome stain as previously described (Patil et al., 2008).

Briefly, smear was fixed in absolute methanol for 10 minutes and allowed to airdry. The air dried smear was stained in modified trichrome stain for 90 minutes and differentiated in 1% acid-alcohol for 3 seconds. Smear was rinsed in two grades of alcohol (95%, 100%), 5 minutes in each. The stained smears were examined microscopically the for spores of microsporidia. The rainy season was defined as the period between the months of April and September while the dry season was between October and March.

Statistical analysis

The data obtained were analyzed using Chi square (X^2) to compare the frequency of data. The odd ratio was calculated for potential risk factor. The software INSTAT (GraphPad Software Inc., La Jolla, CA, USA) was used in all statistical analysis.

RESULTS AND DISCUSSION

Generally, the prevalence of microsporidiosis was significantly associated with season. More episodes of intestinal parasitic infections were observed in the rainy season than the dry season (OR = 1.990; 95% CI = 1.374, 2.882; p = 0.0004). Out of the intestinal parasites observed in this study, only *Microsporidia* was associated with rainy season (Table 1). The least episode of infection was observed in *Entamoeba histolytica* in both rainy and dry seasons (Table 1).

Microsporidia are one of the leading causes of diarrhea in people living with HIV (Weber *et al.*, 1994; Schwartz *et al.*, 1996). With the emergence of AIDS microsporidial diarrhea has gained significance, as it is one of the important causes of morbidity and mortality (Tuli *et al.*, 2008). It has been suggested that seasonal variations in the occurrence of intestinal protozoan infections holds epidemiological significance (Tuli *et al.*, 2008). Hence, this study aims at determining seasonal variations in the occurrence of microsporidiosis.

Of the eleven different types of parasites recovered, only microsporidia species was significantly associated with seasonal variation with higher prevalence in the rainy season.

Bodies of water and animals are known sources of microsporidial infection (Weiss, 2001; Slodkowicz-Kowalska et al., 2006; Cama et al., 2007). It is possible that droppings from wild and domestic animals which may contain microsporidial spores can easily be washed by surface-running water after rainfall into streams and rivers. These bodies of water are used as sources for drinking and domestic use (Akinbo et al., 2010). This will increase the chances of microsporidial infections among immunodeficient persons. This may explain the finding in this study.

The finding of association between microsporidial infection and seasonal variation is of epidemiological importance. This information is important in understanding the transmission, control management of microsporidiosis among HIV-infected patients. It is important to note that only *Ascaris lumbricoides* and hookworm were recovered from non-HIV infected patients and neither was associated with seasonal variations.

Organism	Rainy (%)	Dry (%)	OR	95% CI	P value
E. histolytica	2 (0.8)	3 (1.4)	0.590	0.098, 3.565	0.896
Isospora belli	11 (4.5)	5 (2.3)	2.003	0.685, 5.859	0.300
Microsporidia	61 (24.5)	28 (12.8)	2.250	1.376, 3.677	0.002
A. lumbricoides	39 (15.9)	28 (12.8)	1.285	0.761, 2.170	0.420
Hookworm	18 (7.4)	11 (5.0)	1.492	0.688, 3.234	0.408
S. stercoralis	9 (3.7)	5 (2.3)	1.625	0.536, 4.925	0.553
T. trichiuris	4 (1.6)	7 (3.2)	0.500	0.144, 1.733	0.419
<i>Taenia</i> spp	0	4 (1.8)	0.097	0.005, 1.815	0.104

Table 1: Seasonal trend on the prevalence of intestinal parasitic infections in HIV-infected persons.

In conclusion, this study underscores the association between microsporidiosis and seasonal variations with higher prevalence during the rainy season. A clear understanding of this seasonal dynamics may be helpful in preventing infection.

ACKNOWLEDGEMENT

We acknowledge with thanks the Management of University of Benin Teaching Hospital for permission to carry out this study.

REFERENCES

- Akinbo FO, Okaka CE, Omoregie R. 2010. Prevalence of intestinal parasitic infections among HIV patients in Benin City, Nigeria. *Libyan J Med* 5: 5506 -DOI:10.3402/ljm.v5i0.5506
- Cama, V.A., Pearson, J., Cabrera, L., Pacheco, L., Gilman, R., Meyer, S., Ortega, Y., Xiao, L. 2007. Transmission of Enterocytozoon bieneusi between a child and guinea pigs. *J Clin Microbiol.* 45: 2708-2710
- Cheesbrough M. 1999. Parasitological tests. Cambridge University Press. pp 178-309.
- Conteas C N, Berlin O G, Lariviere M J, Pandhumas SS, Speck CE, Porschen R, Nakaya T. 1998. Examination of the prevalence and seasonal variation of intestinal microsporidiosis in the stools of persons with chronic diarrhea and human immunodeficiency virus infection. Am J Trop Med Hyg. 58(5):559-61
- Cotte L, Rabodonirina M, Chapuis F, Bailly F, Bissuel F, Raynal C, Gelas P, Persaat F, Piens MA, Trepo C. 1999. Waterborne outbreak of intestinal microsporidiosis in persons with and without Human Immunodeficiency Virus infection. *Journal of Infecious Diseases*. 180: 2003-8

Didier E.S. and Weiss L.M. 2006.

Microsporidiosis: current status. *Curr. Opin. Infect. Dis.* 19: 485-492.

- Patil K., De A., Mathur M. 2008. Comparison of weber green and ryan blue modified trichrome staining for the diagnosis of microsporidial spores from stool samples of HIV-positive patients with diarrhea. Indian Journal of Medical Microbiology 26, 407.
- Schwartz D A, Sobottka I, Leitch G J, Cali A, Visvesvara G S. 1996. Pathology of microsporidiosis: emerging parasitic infections in patients with acquired immunodeficiency syndrome. Arch. Pathol. Lab. Med. 120:173–88.
- Slodkowiez-Kowalska A, Graczyk TK, Tamang L, Jedrzejewski S, Nowosad A, Zduniak P, Solarczyk P, Gitouard AS, Majewska AC. 2006. Microsporidium species known to infect humans are present in aquatic birds: Implications for transmission via water. App Environ Microbiol. 72: 4540-4544
- Sokolova O I, Anton V. Demyanov A V, Bowers L C, Didier E L, Yakovlev A V, Sergei O. Skarlato S O, Sokolova Y Y. 2011. Emerging Microsporidian Infections in Russian HIV-Infected Patients. Journal of Clinical Microbiology. 49:2102-2108.
- Tuli L, Gulati A K, Sundar S, Mohapatra T M. 2008. Correlation between CD4 counts of HIV patients and enteric protozoan in different seasons – An experience of the tertiary care hospital in Varanasi (India). *Biomed. Centr. Gastroenterol.* 8:36.
- Tumwine JK, Kekitiiwa A, Nabukeera N, Akiyoshi DE, Buchholt MA, Tzipori S. 2002. Enterocytozoon bieneusi among children with diarrhea attending Mulago Hospital in Uganda. Am J Trop Med Hyg. 67: 299-303
- Van Gool T, Luderhoff E, Nathoo K J, Kiire

CF, Dankert J, Mason PR. 1995. High prevalence of *Enterocytozoon bieneusi* infections among HIV-positive individuals with persistent diarrhea in Harare, Zimbabwe. *Trans R Soc Trop Med Hyg.* 89: 478-80

- Weber R, Bryan R T, Schwartz D A, Owen R L. 1994. Human microsporidial infections. *Clin Microbiol Rev.* 7: 426-461
- Weiss LM. 2001. Microsporidia: emerging pathogenic protists. *Acta Trop.* 78: 89-102.