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Bioscience Research

Print ISSN: 1811-9506 Online ISSN: 2218-3973 Journal by Innovative Scientific Information & Services Network



RESEARCH ARTICLE BIOSCIENCE RESEARCH, 2020 17(4): 4136-4143. OPEN ACCESS

Efficiency of growth cultures of *Bacillus subtilis* with *Luffa cylindrica* used as carbon source

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The Tori (*Luffa cylindrica* L.) also known as Sponge Gourd (SG) is cultivated in tropical and sub-tropical regions of the world. Its fruit is useful as vegetable while rest has been remained non-edible agro-waste material. Present study was aimed to examine cell growth attributes of *Bacillus subtilis* (k1) in sub-merged fermentation cultures supplemented with Tori as carbon source. The fermentation cultures with 12.5 % extracts (v/v) of rotten Tori fruit were maintained i.e. LB_0 [TY-medium: 10.0 g I⁻¹ trypton, 5.0 g I⁻¹ NaCl, 5.0 g I⁻¹ yeast extract in dH₂O (w/v)], LB_1 (¹/₈ LB₀), LB_2 (LB₁ + peels of Tori) and LB_{2a} (LB₁ + peeled Tori) for 18 hrs of incubation. Cell growth and sugar contents were observed significantly higher in LB₂, while reducing sugars and phenolic contents in LB_{2a} culture supernatants. The activities of hydrolytic enzymes like as xylanases and pectinases also observed higher in LB₀ and LB₂ cultures (p≥0.05). Further significant increase in their activities observed in the cultures added with 5.0 m mol⁻³ cadmium (Cd²⁺). In conclusion, Tori are rich with various phyto-metabolites to be the useful agro-nutrient rich fermentation substrate for the production industrial enzymes as well as other secondary metabolites.

Keywords: Bacillus subtilis, sponge gourd, antioxidants, xylanases, pectinases, metal stress.

INTRODUCTION

The Tori (local name) or sponge gourd (Luffa cylindrica L., 2n = 26) member of Cucurbitaceous is cultivated mostly in tropical and subtropical regions (Wu et al. 2016). Its immature fruit is used as vegetable, while fully-grown stuff and whole plant remained unmanaged wastage. Biochemically, sponge gourd is rich with many bioactivators possesses with anti-inflammation, antibacterial, anti-fungal, anti-myocardial properties including anthocyanins (0.5 mg g⁻¹), flavonoids (17.94 mg g⁻¹), glycosides, triterpenoid, saponins, cardiac glycosides, carbohydrates (29.51 %), alkaloids, proteins (33.55 %) fibers (6.47 %), fat (22.17 %), phenolics (20.74 mg g⁻¹), ascorbic acid (1.2 mg g⁻¹) and tannins (37-55%) comparable in concentrations from leaves to seed (Irshad et al. 2010; Osuagwu and Edeoga, 2014; Tripathy et al. 2014). The Tori plant body contains 10 % lignin, 30

% hemi-cellulose and 60 % cellulose (Rowell, 2000). Among the in-organic, *L. cylindrica* is rich with mineral contents including Mg, Na, Ca, K, Cu, Fe, Mn and Zn). This above nutritional potential of Tori is also very important source of vegetable protein, which has remained equally beneficial nutrition for both animal and human (Dairo et al. 2007).

The Tori are rich with growth inducing valuable agro-nutrients and being supportive to maintain invitro cultures (Tanobe et al. 2005). Recent studies reported that it has been used in bioreactor for production of ethanol from raw cassava starch (Roble et al. 2003). This sponge gourd has also been used in the nutrient cultures to prepare hepatocyte cell line of human (Chen et al. 2003). It has good performance in biofilm formation when used as solid state-substrate with maximum capabilities to adsorb nitrifying organic and inorganic agents (de Sousa et al. 2008). Being a suitable natural matrix has been used for biosorption of cadmium (Cd) and lead (Pb) metals successfully (Saeed and Iqbal, 2013: Boudechiche et al. 2016). Such compact and rich with lignocellulosic plants could be the major component in various wastages from household to agriculture including vegetable markets. It may be the staple source of fermentation substrate (Behera et al. 2014).

The physio-chemical conditions including the structural factors of complex agro-waste based substrates hinder the biocatalyst based hydrolysis for their conversion to microbial-fuels. In past decade, a series of chemical based pre-treatment methods have been employed for fermentation biomass preparations which are increasing the cost of fermentation including the 10 % loss of yields from lignocelluloses (Gräsvik et al. 2014; Kucharska et al. 2018). Meanwhile, these loses are increasing the costs of bio-productions. The biodegradation of energy-rich agro-wastes by microorganisms has great economic deal over physio-chemical approaches (Abd-Elsalam and El-Hanafy, 2009; Sánchez, 2009; Bansal et al. 2015; Belal et al. 2018). In agro-wastes, about 60 % of plant biomass is lignocellulose. It is comprised on renewable cellulose and hemicelluloses, which are linked with lignin covalently (Tengerdy and Szakacs, 2003). The Bacillus spp., are diversified fermentation organism that have proven the best producer of many hydrolyzing extra-cellular enzymes including xylanases (Bansal et al. 2014; Mullai et al. 2010), cellulose (Mawadza et al. 2000), lipases (Al-limoun, 2019), pectinases (Kashyap et al. 2003) and strain of bioflocculant production (Goldern et al. 2019) etc.

By keeping in view of aforementioned facts, the present study was planned to utilize the agroindustrial wastes of Tori (*Luffa cylindrica* L.) as fermentation substrate. The *Bacillus subtilis* (k1) was cultured to analyze the potential of substrate on cell growth and other extra-cellular productions including enzymes. These extra-cellular bioproductions ultimately will effect on the decrease of their costs positively.

MATERIALS AND METHODS

Collection of plant materials

For present experiment, agro-waste of Tori or sponge gourd (*Luffa cylindrica* L.) fruit was used as fermentation substrate. It is a well-known fruit and used as vegetable. It was collected from Korai Vegetable and Fruit Market of the city. The collected stuff was further prepared to use in the lab.

Preparation of fermentation substrate

The collected stuff of rotten Tori fruit was washed with running tap- H_2O and dried with toilettowels. The peels of the fruit were peeled-off with fine knife. Both peels and peeled material were used to prepare their filtrates. The 100 g of each were taken in the container of electric-grinder and then 100 ml of d H_2O was added. The mixture was grinded in grinder until homogenous mass of appeared. Both semi-solid plant masses were filtered with muslin-cloth at the room temperature. These fresh-filtrates were used as fermentation substrate.

Preparation of Tori based fermentation cultures

The Tori agro-waste based fermentation cultures were prepared in liquid broth nutrient LB_o (1.0 % Bacto-trypton, 0.5 % yeast extract, 0.5 % NaCl and pH 7.0) medium. Exactly 12.5 % Tori's extracts (filtrate of peels and peeled stuff) sustained low concentrated (LB₁: $\frac{1}{16}$ strength LB_o) medium, while LB₀ medium was considered as positive and LB₁ as nutrient deficit control medium (Table 1). These fermentation cultures were sterilized under standard autoclave-conditions (121°C, 20 min, 15 psi) and it was used when cool down to room temperature.

Preparation of inoculum of fermentation organism

The glycerol stock of *Bacillus subtilis* (k1) was used to prepare for its master-culture. The wireloop was dipped in its stock and inoculated in 2 ml LB_0 broth (Table 1). After over-night incubation at 37°C with 250 rpm shaking (de Vries et al. 2004), its 0.1 ml was sub-cultured in 5 ml LB_0 medium. After 30 min of incubation, it was used as a master culture to inoculate all other culture as shown in table 1.

Harvesting of *Bacillus* fermentation culture

After incubation of bacterial fermentation cultures for 18 hrs under *Bacillus* specific growth conditions (37°C with 250 rpm shaking), their OD600 was taken. After that cultures were harvested and their supernatants were separated with centrifugation for 10 min at 7,000 rpm. It was transferred to aseptic-dark-colored bottles and its pallets were freezed at -20°C. Culture-supernatant were stored at 4°C until next use for different biochemical and enzyme activities.

Biochemical analysis of *Bacillus* culture

The collected culture supernatant was subjected for analysis of various biochemical productions. Like as, total protein contents were measured by taking 1.0 ml culture supernatant in glass test-tube and 2.50 ml alkaline Copper reagent mixed thoroughly. After 10 minutes, 0.25 ml folin-ciocalteu reagent (1:1 dH₂O v/v) was added. By keeping 30 minutes at room temperature, its absorbance was read against blank at OD750 (Lowry et al. 1951). Total sugars were quantified by mixing 0.5 ml supernatant with 2.5 ml H_2SO_4 (conc.) than 0.05 ml phenol (80 %) mixed. After 15 minutes, OD485 was read (Montgomery, 1961). The reducing sugars were also measured in culture supernatant by mixing its 1.0 ml with 1.0 ml 2,6-dinitrosalicylic acid (DNS) in test tube. Mixture was heated for 5 minutes at H₂Oboiling point in H₂O-bath then its OD540 was taken (Miller, 1959). Similarly, phenolics were also measured (Yasoubi et al. 2007) in cultures by taking its 0.2 ml in test tube mixed with 1.0 ml folinciocalteu than 0.8 ml Na₂CO₃ was added. After keeping the mixture at room temperature for 30minutes, OD765 was taken. The flavonoids and antioxidants were also measured in the culture supernatant by following the methods of (Li et al. 2007) and (Djeridane et al. 2006) respectively.

Extra-cellular enzyme activities under cadmium (Cd²⁺) stress

Culture-supernatant of Bacillus subtilis (k1) subjected as crude enzyme for analysis of xylanases and pectinases activities. The xylanases activities determined by taking 1 ml supernatant mixed with 1 ml substrate (0.5 % xylose). After incubation for 15 min at 60°C, 2 ml DNS added then OD540 was noted (Bailey et al. 1992). Similarly, pectinases (Miller, 1959) were assayed by mixing 0.05 ml supernatant as crude enzyme with 1.0 ml substrate [0.25 % pectin in 50 mM citriccitrate buffer (pH 3.5)] than 1.5 ml DNS added (Miller, 1959; Merín et al. 2011). The thermosability of both xylanases and pectinases activities also determined under 5.0 m mol-3 per liter concentration of cadmium (Cd2+) metal ions stresses. These above same reactions were also conducted and incubated at 37°C for 30 min.

Statistical analysis of data

The data of present study was computed for culture significance (ANOVA - analysis of variance and DMR - Duncan's Multiple Range test at 5 % probability level with "COSTAT" package by *CoHort* Software, Berkeley, USA (Snedecor and Cochran, 1967).

RESULTS AND DISCUSSION

Cell growth and effects of carbon sources

The growth of cells and organisms require energy and carbon compounds are rich-sources of essential energy for their movement and reproduction (Hodges, 2010; Jones, 2012). Agrowastes are readily available as ligno-cellulosic rich and economic carbon sources. In present study, effects of Tori or sponge gourd (Luffa cylindrica L.) fruit used as carbon sources on Bacillus subtilis (k1) growth was assessed in the culture harvest taken after 18 hrs of incubation and shown as in table 1. The optimal standard Bacillus spp., growth culture is TY-medium (LB₀: Table 1) with OD600 reached to 1.555±0.029. It was comparable to its cultures maintained on peels of Tori fruit (LB₂) and peeled Tori fruit (LB_{2a}) used as carbon source with OD600 observed to 1.568±0.055 and 1.528±0.081 respectively (Table 2). The glucose is preferred nutrient source for fermentation organisms including B. subtilis (Meyer et al. 2014), while the glucose production ability of the organisms depends on type of secreted hydrolyzing enzymes and available substrate in the fermentation medium (Nigam and Singh, 1995; La Grange et al. 2010; Kampen, 2014).

Table 1: Preparation of agro-waste based fermentation medium for *Bacillus subtilis* (k1) supplemented with agro-wastes of Tori (*Luffa cylindrica* L.) as substrate.

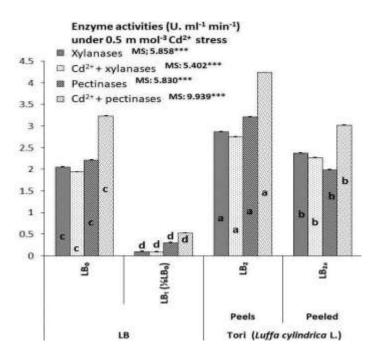
#s.	Medium	Composition of medium		
01	LΒο	TY-medium: Bacto-trypton (1.0 %), NaCl (0.5 %), yeast extract (0.5 %) in distilled-H ₂ O (w/v)		
02	LB1	¼ LB₀ in dH₂O (v/v)		
03	LB ₂	$\label{eq:lb1} \begin{array}{l} LB_1 + 12.5 \text{ ml peels of tori} + 12.5 \\ ml \mbox{ distilled-} H_2O \ (12.5 \ \%, \ v/v) \end{array}$		
04	LB_{2a}	LB ₁ + 12.5 ml peeled tori + 12.5 distilled-H ₂ O (12.5 %, v/v)		

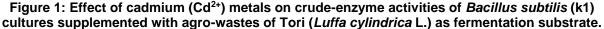
Note: The final concentration of each extract adjusted to 12.5 % in 50 ml final volume of medium. Each *culture maintained with 4 replicates with 50 ml final volume.*

Table 2: Cell growth of *Bacillus subtilis* (k1) and its related extra-cellular attributes in cultures supplemented with agro-wastes of Tori (*Luffa cylindrica* L.) as fermentation substrate.

<i>#</i> ~	Channa taniatian	LB		Tori (<i>Luffa cylindrica</i> L.)		F-
#s.	Characteristics	LB_{o}	LB₁ (½LB₀)	LB_2	LB_{2a}	significance
01.	Cell growth (OD ₆₀₀)	1.555±0.029 ^b	0.260±0.009°	1.568±0.055 ^a	1.528±0.081 ^{bc}	159.5***
02.	Total sugars (mg ml⁻¹)	22.18±0.134 ^b	8.636±0.290°	30.06±0.087 ^a	29.53±0.258 ^a	2261***
03.	Reducing sugars (mg ml⁻¹)	6.058±0.116 ^c	2.927±0.241 ^d	8.801±0.116 ^{ab}	9.0230.1385 ^a	80.32***
04.	Total proteins (mg ml ⁻¹)	7.299±0.059°	2.551±0.220 ^d	9.319±0.234 ^a	8.669±0.129 ^b	302.3***
05.	Ascorbic acid (mg ml ⁻¹)	3.735±0.112℃	1.990±0.064 ^d	4.855±0.071 ^a	4.800±0.073 ^a	267.1***
06.	Proline (mg ml ⁻¹)	0.057±0.002 ^a	0.026±0.001 ^b	0.055±0.001 ^a	0.057±0.002 ^a	120.3***
07.	Phenolics (mg ml ⁻¹)	2.833±0.160 ^b	2.833±0.132 ^b	5.778±0.143 ^a	5.833±0.289 ^a	80.08***
08.	Flavonoids (mg ml ⁻¹)	0.341±0.018 ^b	0.176±0.010 ^c	0.474 ± 0.005^{a}	0.498±0.011ª	150.4***
09.	Antioxidants (mg ml ⁻¹)	0.912±0.028 ^b	0.343±0.021°	1.847±0.022 ^a	1.822±0.020 ^a	1034.6***

The collected data is mean of 4-replicates of all treatments or cultures [mean \pm SE (n = 4)]. The Duncan's Multiple Range test was performed at p \geq 0.05 level of ANOVA significance.





Productions of various extra-cellular metabolites

With rapid increase in cell multiplication rates on different cultures supported with control to agrowaste based carbon sources, various extracellular metabolites were increased. Like as total sugars and reducing sugars were higher in LB_{2a} culture supernatants which were harvested after

18 hrs of incubation than the control cultures

(Table 1 & 2). Similarly other metabolites, total protein contents and antioxidants of fermented substrates were higher in LB₂ and LB_{2a} cultures. The *Bacillus subtilis* is serving as single cell protein secretor. In other studies it has been reported that protein contents increases when ligno-cellulosic rich agro-wastes are used as fermentation substrate (Ofuya and Nwajiuba, 1990; Shabeb et al. 2010; Zerdani et al. 2004). The other antioxidants also observed with differential concentrations among the culture supernatants.

Maximum proline and ascorbates were observed among the agro-wasted based cultures LB_{2a} and LB₂ supernatants respectively ($p \ge 0.05$), while phenolics and flavonoids were higher in LB_{2a} cultures (Table 2). With increase in culture time, the production of metabolites in the cultures increases, which lead to develop specific detrimental effects on culture growth as well as extra-cellular bioproductions (Misawa, 1994; Sravanthi et al. 2016). For mitigation of the developed stress, the cell triggers the endogenous free proline and ascorbate accumulation in different organisms (Wood et al. 2001; Shivanand and Mugeraya, 2011).

Extra-cellular enzymes productions

The results of xylanases and pectinases activities in the control as well as Tori agro-waste based submerged fermentation (SSF) culture supernatants of B. subtilis (k1), which were harvested after 18 hrs of incubation (Fig 1). The maximum pectinases and xylanases activities among the culture supernatants were observed in LB₂ (Tori-peels) cultures than other LB_o (TYmedium) and LB2a (peeled-Tori). The availability of xylan and pectin in the culture triggers the production of their respective hydrolyzing enzymes (xylanases and pectinases) like as the richness of cellulases activities in cellulose rich extracts. Even hemicellulose are barrier for cellulases access to cellulose that's why its hydrolysis is essential for efficient agro-waste saccharification (Zimbardi et al. 2013). The above fact is due to that cultured organism is producing all enzymes required to digest the biomass used for its growth (Kumar et al. 2008; Znameroski et al. 2012).

Cadmium (Cd²⁺) stresses on enzyme activities

According to the figure 1, it shows the activities of xylanases and pectinases in agro-biomass based culture supernatants of B. subtilis (k1), which were harvested after 18 hrs of incubation. With the application of 0.5 m mol⁻³ CdCl₂ (Cd²⁺) stress, its significant effect on enzyme activities was observed (Fig 1). The xylanase activities were decreased, while pectinases showed increase in its activities in all culture supernatants. Such inhibitions as well as enhancements are adopted naturally by the indigenous microorganism for biotransformation of toxic metals (Benammar et al. 2015; Voica et al. 2016). It is specific required industrial features for attraction of commerce community. Among the saline bacteria, it is widespread character (Nieto Gutiérrez et al. 1989; Cánovas et al. 2000; Amoozegar et al. 2005). For xylanases, Cd2+ has shown inhibitory effect, while

remains enhancer for pectinases activities. It means that cadmium metal ions (Cd²⁺) are playing vital role to keep active confirmation to stimulate pectinases activities (Beg and Gupta, 2003).

The main applications of hydrolytic enzymes are to utilize complex agro-waste biomass conversion into simple absorbable carbon molecules by the microbes. Various groups have studied in order to test saccharification potential of celluloses for rice straw (Ball and McCarthy, 1988) or other cellulose rich organic biomasses (Prasad et al. 2012). In the last decade, various typed agrobiomass have been utilized for sustainable productions of secondary metabolites enzymes and biofuels and chemicals (Puri and Abraham, 2016; Voloshin et al. 2016; Kumar et al. 2017; Vellaichamy and Saxena, 2020). At the moment, present study is carried out to evaluate the potential of cellulosic extract of Tori plant fruit with Bacillus subtilis (k1) for bio-production of metabolites and saccharification enzymes like as xylanases and pectinases.

CONCLUSION

The in-vitro cellular productions are considered economic, natural and safe than via chemical methods with excess energy. The utilization of low cost lingo-cellulosic rich agro-wastes could fulfill the economic energy needs. Tori-wastes are full with carbon and nitrogen sources for Bacillus subtilis, which has significant role in industrial production including various enzymes. Like as the cultures supplemented with Tori's wastes (LB2 and LB_{2a} cultures) shown comparative (p≥0.05) cell growth, production of metabolites and important industrial enzymes. Present work could be helpful in future for selection of energy-rich part of plant wastes for the production of maximum extracellular metabolites and its subsequent fermentation organism.

CONFLICT OF INTEREST

The authors declared that present study was performed in absence of any conflict of interest.

ACKNOWLEGEMENT

We, the authors like to be highly thankful to University of Sindh, Jamshoro for potential provision of financial support for the completion of present study. Authors are also grateful to technical and non-technical supporting staff of the laboratories of the respective institute for their timely co-operations.

AUTHOR CONTRIBUTIONS

Author 1 and 2 has carried out the present research work, while Author 2 designed the objectives and guided throughout the study. Author 3 tabulated the data and statistical analysis. Author 4 & 5 have pre-submitted review this present manuscript critically.

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REFERENCES

- Abd-Elsalam H.E. and El-Hanafy A.A., 2009. Lignin biodegradation with ligninolytic bacterial strain and comparison of *Bacillus subtilis* and *Bacillus* sp. isolated from Egyptian soil. Amr. Eur. J. Agric. Environ. Sci. 5(1): 39-44.
- Al-limoun M.O., 2019. Potential bioremediation of waste corn oil by extracellular lipase enzyme from *Monascus purpureus* W7. Biosci. Res. 16(4): 3508-3522.
- Amoozegar M. Hamedi J. Dadashipour M. and Shariatpanahi S., 2005. Effect of salinity on the tolerance to toxic metals and oxyanions in native moderately halophilic spore-forming *Bacilli*. World J. Microb. Biotechnol. 21(6-7): 1237-1243.
- Bailey M.J. Biely P. and Poutanen K., 1992. Interlaboratory testing of methods for assay of xylanase activity. J. Biotechnol. 23(3): 257-270.
- Ball A.S. and McCarthy A.J., 1988. Sacchariiication of straw by actinomycete enzymes. Microbiol. 134(8): 2139-2147.
- Bansal N. Soni R. Janveja C. and Soni S.K. 2015. Production of xylanase-cellulase complex by *Bacillus subtilis* NS7 for the biodegradation of agro-waste residues. Lignocellulosic. 1(3): 196-209.
- Bansal N. Soni R. Janveja C. and Soni S.K., 2014. Production of xylanase-cellulase complex by *Bacillus subtilis* NS7 for the biodegradation of agro waste residues. Appl. Biochem. Biotechnol. 172(1): 141-156.
- Beg Q.K. and Gupta R., 2003. Purification and characterization of an oxidation-stable, thioldependent serine alkaline protease from

Bacillus mojavensis. Enzyme Microb. Technol. 32(2): 294-304.

- Behera S. Arora R. Nandhagopal N. and Kumar S., 2014. Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. Renew. Sustain. Energy Rev. 36: 91-106.
- Belal E.-S.B. Shalaby M.E. El-Gremi S.M. and Gad W.A., 2018. Biodegradation of organochlorine pesticides by *Paenibacillus* sp. Strain. Environ. Engr. Sci. 35(11): 1194-1205.
- Benammar L. Menasria T. Ayachi A. and Benounis M., 2015. Phosphate removal using aerobic bacterial consortium and pure cultures isolated from activated sludge. Proc. Safety Environ. Prot. 95: 237-246.
- Boudechiche N. Mokaddem H. Sadaoui Z. and Trari M., 2016. Biosorption of cationic dye from aqueous solutions onto lignocellulosic biomass (*Luffa cylindrica*): characterization, equilibrium, kinetic and thermodynamic studies. Int. J. Indut. Chem. 7(2): 167-180.
- Cánovas D. Vargas C. Kneip S. Morón M.A.-J. Ventosa A. Bremer E. and Nieto J.N.J., (2000). Genes for the synthesis of the osmoprotectant glycine betaine from choline in the moderately halophilic bacterium *Halomonas elongata* DSM. Microbiol. 146(2): 455-463.
- Chen J.P. Yu S.C. Hsu B.R.S. Fu S.H. and Liu H.S. 2003. Loofa sponge as a scaffold for the culture of human hepatocyte cell line. Biotechnol. Prog. 19(2): 522-527.
- Dairo F. Aye P. and Oluwasola T., 2007. Some functional properties of loofah gourd (*Luffa cylindrica* L.) seed. J. Food Agric. Environ. 5(1): 97-101.
- de Sousa J.T. Henrique I.N. Oliveira R. Lopes W.S. and Leite V.D. 2008. Nitrification in a submerged attached growth bioreactor using *Luffa cylindrica* as solid substrate. Afr. J. Biotechnol. 7(15): 2702-2706.
- de Vries Y.P. Hornstra L.M. de Vos W.M. and Abee T., 2004. Growth and sporulation of *Bacillus cereus* ATCC 14579 under defined conditions: temporal expression of genes for key sigma factors. Appl. Environ. Microbiol. 70(4): 2514-2519.
- Djeridane A. Yousfi M. Nadjemi B. Boutassouna D. Stocker P. and Vidal N., (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. Food Chem. 97(4): 654-660.
- Goldern Z. Mthembu N.S.,Gasa N.L. Kotze B.A. Simonis J.J. Evelyn M. and Rajasekhar P.V.S. 2019. Isolation, identification and

characterization of a bioflocculant producing strain, *Bacillus spp.* KC782848.1, from umlalazi catchment, Mtunzini, kwaZulu-Natal. Biosci. Res. 16(4): 3664-3685.

- Gräsvik J. Winestrand S. Normark M. Jönsson L.J. and Mikkola J.-P., 2014. Evaluation of four ionic liquids for pretreatment of lignocellulosic biomass. BMC Biotechnol. 14(1): 34.
- Hodges S.C., 2010. Soil fertility basics. Soil Science Extension, North Carolina State University.
- Irshad M. Ahmad I. Goel H. and Rizvi M.M.A., 2010. Phytochemical screening and high performance TLC analysis of some cucurbits. Res. J. Phytochem. 4: 242-247.
- Jones J.J.B., 2012. Plant nutrition and soil fertility manual: CRC press.
- Kampen W.H., 2014. Nutritional requirements in fermentation processes *Fermentation and biochemical engineering handbook.* pp 37-57.
- Kashyap D.R. Soni S.K. and Tewari R., 2003. Enhanced production of pectinase by *Bacillus* sp. DT7 using solid state fermentation. Biores. Technol. 88(3): 251-254.
- Kucharska K. Rybarczyk P. Hołowacz I. Łukajtis R. Glinka M. and Kamiński M., 2018. Pretreatment of lignocellulosic materials as substrates for fermentation processes. Molecules. 23(11): 2937.
- Kumar R. Singh S. and Singh O.V., 2008. Bioconversion of lignocellulosic biomass: biochemical and molecular perspectives. J. Indust. Microbiol. Biotechnol. 35(5): 377-391.
- Kumar S.J. Gujjala L.K.S. Dash A. Talukdar B. and Banerjee R., 2017. Biodiesel production from lignocellulosic biomass using oleaginous microbes *Lignocellulosic Biomass Production and Industrial Applications,* pp 65-92. Wiley Online Library.
- La Grange D.C. Den Haan R. and Van Zyl W.H., 2010. Engineering cellulolytic ability into bioprocessing organisms. Appl. Microbiol. Biotechnol. 87(4): 1195-1208.
- Li W. Gao Y. and Zhao J., 2007. Phenolic, Flavonoid and Lutein Ester Content and Antioxidant Activity of Eleven Cultivars of Chinese Marigold. J. Agricult. Food Chem. 55(21): 8478-8484.
- Lowry O. Rosebrough N. Farr A. and Randall R., 1951. Protein measurement with the Folin phenol reagent. J. Biol. Chem. 193: 265-275.
- Mawadza C. Hatti-Kaul R. Zvauya R. and Mattiasson B., 2000. Purification and characterization of cellulases produced by two *Bacillus* strains. J. Biotechnol. 83(3): 177-

187.

- Merín M.G. Mendoza L.M. Farías M.E. and De Ambrosini V.I.M., 2011. Isolation and selection of yeasts from wine grape ecosystem secreting cold-active pectinolytic activity. Int. J. Food Microbiol. 147(2): 144-148.
- Meyer H. Weidmann H. Mäder U. Hecker M. Völker U. and Lalk M., 2014. A time resolved metabolomics study: the influence of different carbon sources during growth and starvation of *Bacillus subtilis*. Mol. BioSyst. 10(7): 1812-1823.
- Miller G.L., 1959. Use of dinitrosalicylic acid reagent for determination of reducing sugar. Analyt. Chem. 31(3): 426-428.
- Misawa M., 1994. Plant tissue culture: an alternative for production of useful metabolites: Food & Agriculture Org.
- Montgomery R., 1961. Further studies of the phenolsulfuric acid reagent for carbohydrates. Biochim. Biophys. Acta. 48: 591-593.
- Mullai P. Fathima N.S.A. and Rene E.R., 2010. Statistical analysis of main and interaction effects to optimize xylanase production under submerged cultivation conditions. J. Agric. Sci. 2(1): 144-153.
- Nieto G.J.J. Fernández C.R. Márquez M.M.D.C. Ventosa U.A. Quesada E. and Ruiz B.F., 1989. Survey of metal tolerance in moderately halophilic *Eubacteria*. Appl. Environment. Microbiol. 55: 2385-2390.
- Nigam P. Singh D., 1995. Enzyme and microbial systems involved in starch processing. Enzyme Microb. Technol. 17(9): 770-778.
- Ofuya C. and Nwajiuba C., 1990. Microbial degradation and utilization of cassava peel. World J. Microbiol. Biotechnol. 6(2): 144-148.
- Osuagwu A. and Edeoga H., 2014. Nutritional properties of the leaf, seed and pericarp of the fruit of four *Cucurbitaceae* species from South-East Nigeria. IOSR J. Agric. Vet. Sci. 7(9): 41-44.
- Prasad P. Bedi S. and Singh T., 2012. In vitro cellulose rich organic material degradation by cellulolytic *Streptomyces albospinus* (MTCC 8768). Malay. J. Microbiol. 8(3): 164-169.
- Puri M. and Abraham R., 2016. Strategies to Enhance Enzyme Activity for Industrial Processes in Managing Agro-Industrial Waste Agro-Industrial Wastes as Feedstock for Enzyme Production, pp 299-312. Elsevier.
- Roble N. Ogbonna J. and Tanaka H., 2003. A novel circulating loop bioreactor with cells immobilized in loofa (*Luffa cylindrica*) sponge

for the bioconversion of raw cassava starch to ethanol. Appl. Microbiol. Biotechnol. 60(6): 671-678.

- Rowell R.M., 2000. Characterization and factors effecting fiber properties. Natural polymers and agrofibers based composites. Sãn Carlos, Brazil,
- Saeed A. and Iqbal M., 2013. Loofa (*Luffa cylindrica*) sponge: Review of development of the biomatrix as a tool for biotechnological applications. Biotechnol. Prog. 29(3): 573-600.
- Sánchez C., 2009. Lignocellulosic residues: Biodegradation and bioconversion by fungi. Biotechnol. Adv. 27(2): 185-194.
- Shabeb M.S. Younis M.A. Hezayen F.F. and Nour-Eldein M.A., 2010. Production of cellulase in low-cost medium by *Bacillus subtilis* KO strain. World Appl. Sci. J. 8(1): 35-42.
- Shivanand P. and Mugeraya G., 2011. Halophilic bacteria and their compatible solutes– osmoregulation and potential applications. Curr. Sci. 100(10): 1516-1521.
- Snedecor G.W. and Cochran W.G., 1967. Statistical Methods, 6th Edn, pp 129-131. Ames. *Iowa, USA, Lowa State University Press.*.
- Sravanthi M. Mohan G.K. Suryakala G. Rani M.S. and Shanker K., 2016. Plant tissue culture: An alternative for production of useful secondary metabolites. J. Pharmacogn. Phytochem. 5(4): 269-272.
- Tanobe, V. O., Sydenstricker, T. H., Munaro, M., and Amico, S. C. (2005). A comprehensive characterization of chemically treated Brazilian sponge-gourds (*Luffa cylindrica*). Polym. Test. 24(4), 474-482.
- Tengerdy R. and Szakacs G., 2003. Bioconversion of lignocellulose in solid substrate fermentation. Biochem. Engineer. J. 13(2-3): 169-179.
- Tripathy P.K. Kumar S. and Jena P.K., 2014. Nutritional and medicinal values of selected wild cucurbits available in similipal biosphere reserve forest, odisha. Int. J. Pharm. Sci Res. 5(12): 5430-5437.
- Vellaichamy M. and Saxena A.K., 2020. Fermentation technology: A viable tool for bioconversion of lignocellulosic biomass into value-added products. Int. J. Curr. Microbiol. App. Sci. 9(7): 1747-1762.
- Voica D.M. Bartha L. Banciu H.L. and Oren A., 2016. Heavy metal resistance in halophilic Bacteria and *Archaea*. FEMS microbiology letters, 363(14):146.

- Voloshin R.A. Rodionova M.V. Zharmukhamedov S.K. Veziroglu T.N. and Allakhverdiev S.I., 2016. Biofuel production from plant and algal biomass. Int. J. Hyd. Energy. 41(39): 17257-17273.
- Wood J.M. Bremer E. Csonka L.N. Kraemer R. Poolman B. van der Heide T. and Smith L.T., 2001. Osmosensing and osmoregulatory compatible solute accumulation by bacteria. *Comparative Biochemistry and Physiology* (A): Mol. Integ. Physiol. 130(3): 437-460.
- Wu H. He X. Gong H. Luo S. Li M. Chen J. Zhang C. Yu T. Huang W. and Luo J., 2016. Genetic linkage map construction and QTL analysis of two interspecific reproductive isolation traits in sponge gourd. Front. Plant Sci. 209(3): 1067-1082.
- Yasoubi P. Barzegar M. Sahari M.A. and Azizi M., 2007. Total phenolic contents and antioxidant activity of pomegranate (*Punica granatum* L.) peel extracts. J. Agric. Sci. Technol. 9: 35-42
- Zerdani I. Faid M. and Malki A., 2004. Feather wastes digestion by new isolated strains *Bacillus* sp. in Morocco. Afr. J. Biotechnol. 3(1): 67-70.
- Zimbardi A.L. Sehn C. Meleiro L.P. Souza F.H. Masui D.C. Nozawa M.S. Guimarães L.H, Jorge J.A, and Furriel R.P., 2013. Optimization of β -glucosidase, β -xylosidase and xylanase production by *Colletotrichum graminicola* under solid-state fermentation and application in raw sugarcane trash saccharification. Int. J. Mol. Sci. 14(2): 2875-2902.
- Znameroski E.A. Coradetti S.T. Roche C.M. Tsai J.C. lavarone A.T. Cate J.H. and Glass N.L., 2012. Induction of lignocellulose-degrading enzymes in *Neurospora crassa* by cellodextrins. Proc. Natl. Acad. Sci. 109(16): 6012-6017.